

Policy #:	602 (PLH-602-02)	Effective Date:	9/30/2004	Reviewed Date:	2/4/2011
Subject:	COLLECTION OF CEREBROSPINAL FLUID & OTHER STERILE BODY FLUIDS FOR CULTURE				
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COLLECTION OF CEREBROSPINAL FLUID & OTHER STERILE BODY FLUIDS FOR CULTURE

Lumbar puncture:

1. Suggested volumes of CSF are 1, 2, and 3 ml for routine, fungal, and mycobacterial cultures, respectively. Optimal recovery of Fungal and mycobacterial cultures obtained from 4 mls. of CSF.
2. Clean the puncture site with antiseptic solution and alcohol before needle insertion to prevent introduction of infection.
3. Slowly drain the CSF into the sterile leakproof tubes. Three tubes are generally required for microbiology, hematology, and chemistry testing. The second tube drawn will generally go to microbiology, and the last tube drawn will generally go to hematology. (In traumatic taps, the CSF will often clear as the later tubes are collected.) *Note: always send the most turbid tube to microbiology.*

Brain abscess:

1. Most brain abscesses will grow anaerobic bacteria and should be collected and transported under anaerobic conditions.
2. Contact the microbiology laboratory prior to specimen collection in order to obtain an appropriate transport system.
3. Aspirated material from a lesion provides a better specimen than swabs of the lesion.
4. Transport the specimen without delay to the microbiology laboratory for immediate processing.

Central Nervous System (CNS) Biopsy Samples:

CNS biopsy obtained from the lesion at surgery should be sent to the microbiology laboratory in an anaerobic transport system. Swabs are inappropriate collection devices for anaerobes. *Do not add formalin.*

Other Sterile body fluids excluding CSF, urine, and blood:

1. Clean the needle puncture site with alcohol and disinfect it with an iodine solution (1 to 2% tincture of iodine or a "10% solution of povidone-iodine [1% free iodine]) to prevent introduction of infection. (If tincture of iodine is used, remove with 70% ethanol after the procedure to avoid burn.)

2. The physician will aseptically perform percutaneous aspiration to obtain pleural, pericardial, peritoneal, or synovial fluids.
3. Expel air bubbles from the syringe, and immediately inject the specimen into an anaerobic transport system or send the specimen in the syringe without the needle, the collection bag or a sterile screw-cap container.